



Temperature-dependent vitamin D signaling regulates developmental trajectory associated with diapause in an annual killifish

Amie L. T. Romney^{a,b}, Erin M. Davis^a, Meranda M. Corona^a, Josiah T. Wagner^c, and Jason E. Podrabsky^{a,1}

^aDepartment of Biology, Portland State University, Portland, OR 97207; ^bSchool of Veterinary Medicine, Anatomy, Physiology, and Cell Biology, University of California, Davis, CA 95616; and ^cKnight Cancer Early Detection Advanced Research Center, Oregon Health and Science University, Portland, OR 97201

Edited by David M. Kingsley, Stanford University, Stanford, CA, and approved October 10, 2018 (received for review March 16, 2018)

The mechanisms that integrate environmental signals into developmental programs remain largely uncharacterized. Nuclear receptors (NRs) are ligand-regulated transcription factors that orchestrate the expression of complex phenotypes. The vitamin D receptor (VDR) is an NR activated by $1\alpha,25$ -dihydroxyvitamin D₃ [$1,25(\text{OH})_2\text{D}_3$], a hormone derived from 7-dehydrocholesterol (7-DHC). VDR signaling is best known for regulating calcium homeostasis in mammals, but recent evidence suggests a diversity of uncharacterized roles. In response to incubation temperature, embryos of the annual killifish *Austrofundulus limnaeus* can develop along two alternative trajectories: active development and diapause. These trajectories diverge early in development, from a biochemical, morphological, and physiological perspective. We manipulated incubation temperature to induce the two trajectories and profiled changes in gene expression using RNA sequencing and weighted gene coexpression network analysis. We report that transcripts involved in $1,25(\text{OH})_2\text{D}_3$ synthesis and signaling are expressed in a trajectory-specific manner. Furthermore, exposure of embryos to vitamin D₃ analogs and $\Delta 4$ -dafachronic acid directs continuous development under diapause-inducing conditions. Conversely, blocking synthesis of $1,25(\text{OH})_2\text{D}_3$ induces diapause in *A. limnaeus* and a diapause-like state in zebrafish, suggesting vitamin D signaling is critical for normal vertebrate development. These data support vitamin D signaling as a molecular pathway that can regulate developmental trajectory and metabolic dormancy in a vertebrate. Interestingly, the VDR is homologous to the *daf-12* and *ecdysone* NRs that regulate dormancy in *Caenorhabditis elegans* and *Drosophila*. We suggest that 7-DHC-derived hormones and their associated NRs represent a conserved pathway for the integration of environmental information into developmental programs associated with life history transitions in animals.

nuclear receptors | life history | dormancy | phenotypic plasticity

There is a growing appreciation for the importance of environmental exposures during development in the determination of a wide range of complex phenotypes beyond classic examples of phenotypic plasticity (e.g., fetal programming in mammalian embryos). Despite their profound influence on form and function, the mechanisms by which phenotypes are modified by environmental factors are largely undefined and unexplored.

The annual killifish *Austrofundulus limnaeus* survives in ephemeral ponds by producing stress-tolerant embryos that survive for months encased in drying mud. Embryos survive by entering into three distinct stages of developmental arrest termed diapause I, II, and III (1–3). Diapause is a state of profound metabolic depression and extreme stress tolerance; diapause II embryos can survive for months without access to oxygen or liquid water (2, 4). Entrance into diapause II (diapause from here forward) is the result of an alternative developmental trajectory that is physiologically, morphologically, and biochemically unique compared with “escape” embryos that develop continuously until hatching or entrance into diapause III (5, 6). Developmental phenotype can be influenced by maternal provisioning and by the embryonic environment. Exposure to increased temperature (30 °C compared with 20 °C to

25 °C) and light promotes the escape trajectory (6, 7). Biochemically, the trajectories diverge early in development, before the formation of the embryonic axis, as evidenced by reduced insulin-like growth factor (IGF) expression in diapause-bound embryos (8). A critical window of development where increased temperature causes irreversible commitment to the escape trajectory occurs during early somitogenesis in embryos possessing 10 to 20 pairs of somites. Importantly, these temperatures are within the bounds experienced by embryos under field conditions. Thus, embryos of annual killifishes offer an unprecedented opportunity to study genome–environment interactions during vertebrate development in an ecologically and evolutionarily relevant context (1, 9, 10).

Here we report that vitamin D₃ synthesis and signaling promotes active development in fish embryos, and inhibition of vitamin D₃ synthesis leads to developmental arrest. In this paper we describe vitamin D₃ signaling as a pathway that regulates developmental trajectory and metabolic dormancy, and allows for the integration of environmental information into the developmental program.

Results

Key Gene Regulatory Networks Are Unique to Each Developmental Trajectory. General results for the RNA sequencing (RNAseq) analysis are provided in *SI Appendix*, Fig. S1 and Table S1 and

Significance

Here we describe a molecular pathway that regulates dormancy in a vertebrate and highlights a mechanism that integrates environmental cues into a developmental program that has clear ecological and evolutionary significance. Further, we provide compelling evidence that vitamin D signaling is critical for normal vertebrate development and can induce a diapause-like arrest in embryos of zebrafish. The vitamin D receptor is homologous to nuclear receptors (NRs) that regulate dormancy in *Caenorhabditis elegans* and *Drosophila*. This conservation of function suggests a conserved role for hormones derived from 7-DHC and their associated NRs in the control of metabolic dormancy and major life history transitions in animals.

Author contributions: A.L.T.R., E.M.D., J.T.W., and J.E.P. designed research; A.L.T.R., E.M.D., M.M.C., J.T.W., and J.E.P. performed research; A.L.T.R., E.M.D., M.M.C., J.T.W., and J.E.P. analyzed data; and A.L.T.R. and J.E.P. wrote the paper.

The authors declare no conflict of interest.

This article is a PNAS Direct Submission.

Published under the PNAS license.

Data deposition: The original data sets are available via the National Center for Biotechnology Information's (NCBI) Sequence Read Archive, <https://www.ncbi.nlm.nih.gov/sra> (accession nos. SRX1032655, SRX1032657, SRX1032658, SRX1032664–SRX1032702; BioProject ID PRJNA272154).

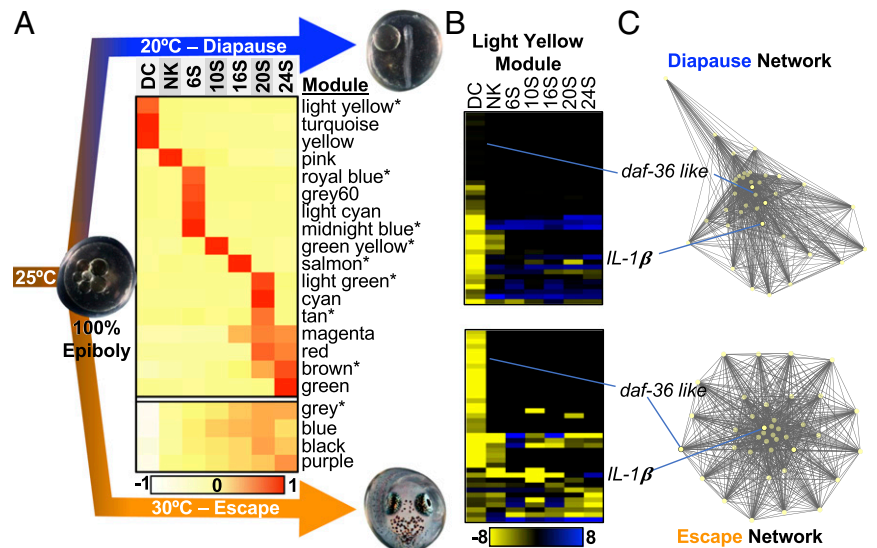
See Commentary on page 12553.

¹To whom correspondence should be addressed. Email: jpod@pdx.edu.

This article contains supporting information online at www.pnas.org/lookup/suppl/doi:10.1073/pnas.1804590115/-DCSupplemental.

Published online November 16, 2018.

Fig. 1. WGCNA of RNAseq data in embryos of *A. limnaeus* developing along alternative developmental trajectories. To identify trajectory-specific gene expression patterns, embryos were reared 20 °C or 30 °C to induce the diapause or escape trajectories, respectively. Embryos ($n = 3$ groups of 20 embryos) were sampled at six morphological stages that bracket the temperature-sensitive window where developmental trajectory is determined: DC, dispersed cell stage; NK, neural keel stage; 6S, 6-somite embryo; 10S, 10-somite embryo; 16S, 16-somite embryo; 20S, 20-somite embryo; 24S, 24-somite embryo. (A) Heat map of module eigengene (based on weighted average of module gene expression) correlations to each stage of development along the diapause trajectory (positive is red, and negative is white). (Upper) Modules are significantly associated ($r > 0.50$; P value $< 10^{-2}$) with a developmental stage. (Lower) Modules are not significantly associated. Modules with asterisks are not preserved between diapause and escape networks (Z_{summary} statistic < 10). (B) Heat maps of expression level reported as fragments per kilobase of transcript per million mapped reads (FPKM) for the light yellow module that contains a *daf-36*-like cholesterol desaturase (LOC106533739) and *IL-1 β* as hub genes. (C) Cytoscape (version 3.5.1) visualization of the light yellow module, with yellow circles representing the genes and the connecting lines representing similarity strengths (thickness) (44), based on topological overlap determined by WGCNA.



Datasets S1 and S2. Weighted gene coexpression network analysis (WGCNA) was used to construct 21 gene coexpression modules from the more than 16,000 transcripts expressed in embryos developing along the diapause trajectory (11) (SI Appendix, Fig. S2). Network topologies of 12 modules were preserved in the escape trajectory and likely represent conserved transcriptional pathways critical to normal development (11, 12). We focused on the remaining nine modules that represent diapause- and escape-specific networks. Based on a measure of intramodular connectivity (kME), we identified 3,034 intramodular “hub” genes ($kME > 0.90$, $P < 10^{-8}$; Dataset S3) in the diapause trajectory as a means to discover key regulators of each module (11).

Notably, within 24 h of temperature transfer [dispersed cell (DC) stage embryos], the transcriptomes of the two trajectories diverged. One of the three WGCNA modules associated with this stage (light yellow) was not preserved in the two trajectories ($Z_{\text{summary}} = 6.2$). This module consists of 39 low-abundance transcripts (Fig. 1B) with two hub genes of particular interest: interleukin-1 β (*IL-1 β* , $kME = 0.97$, $P < 10^{-12}$) and a *daf-36*-like cholesterol desaturase ($kME = 0.98$, $P < 10^{-14}$). These transcripts are escape trajectory-specific with low or undetectable expression in diapause-bound embryos, and there is a dramatic restructuring of the coexpression networks between the two trajectories (Fig. 1C). *IL-1 β* is a potent cytokine with unknown roles in fish development (13), but can regulate IGF signaling in mammals (14). The *daf-36*-like transcript encodes an enzyme that catalyzes the production of 7-dehydrocholesterol (7-DHC), the first step in the synthesis of dafachronic acid hormones that regulate entrance into dauer dormancy in *Caenorhabditis elegans* through the *daf-12* nuclear receptor (NR) (15).

Many Dauer Pathway Genes Are Differentially Expressed in Escape- and Diapause-Bound Embryos. There are four signaling pathways that are known to regulate dauer dormancy in *C. elegans* (16), and key genes in each of these pathways are differentially expressed in *A. limnaeus* embryos developing along the two alternate trajectories (SI Appendix, Fig. S3). Some of the earliest differences in expression are observed in homologs to genes involved in sensory ciliary signaling, insulin-like signaling, and *daf-12* hormonal signaling. Given the identification of the *daf-36*-like transcript as a hub gene and possible regulator of

phenotype, and the key importance of *daf-12* signaling for regulating dauer dormancy in *C. elegans*, we decided to focus further on this pathway.

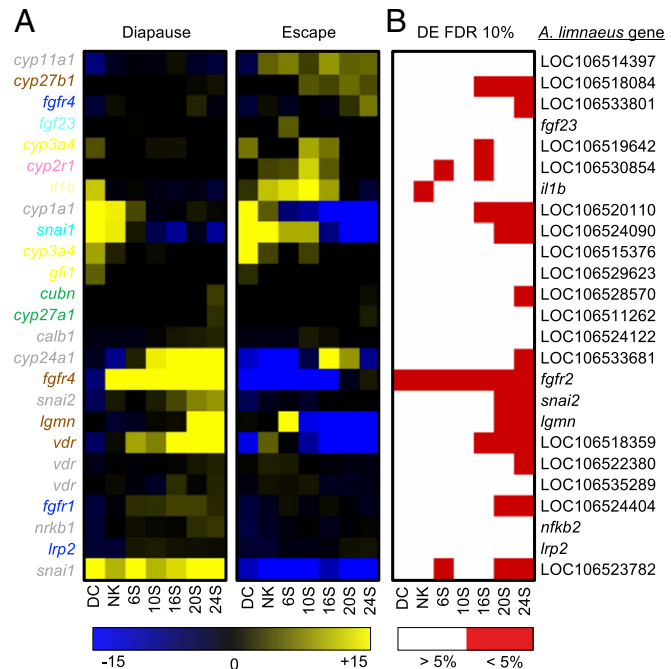


Fig. 2. Vitamin D₃ signaling gene transcripts in *A. limnaeus*. (A) Heat maps of expression levels (FPKM) for vitamin D₃ signaling genes in each developmental phenotype. Mammalian homologs are identified by gene symbol on the left. Text color for each gene symbol represents WGCNA module origin. Clustering was accomplished using uncentered Pearson correlation with complete linkage in Gene Cluster 3.0 (45, 46). Heat maps were generated with Java Treeview 1.1.6r4 using median-centered FPKM values (47). (B) Heat map showing significant FDR adjusted P values (FDR = 10%, $P < 0.05$, calculated in DESeq2) for transcripts differentially expressed in the two phenotypes; DC, dispersed cell stage; NK, neural keel stage; 6S to 24S indicates the number of somite pairs in each subsequent stage.

mammals and with patterns of *daf-12* gene expression in *C. elegans* (24).

The genomic activity of VDR is mediated by the binding of VDR homodimers, or heterodimers with a variety of other NRs, to vitamin D₃ response elements in gene promoters. A rich diversity of NRs is expressed during early development in *A. limnaeus*, many of which exhibit trajectory-specific expression patterns (SI Appendix, Fig. S4). The most common partner for VDR is the retinoid X receptor (25). However, partnering with thyroid hormone receptor can alter the DNA binding affinity and transcriptional activation efficiency of VDR (26). The VDR can also partner with a number of transcriptional repressors and chromatin-modifying complexes that determine complex context-dependent transcriptional networks (25). Thus, we hypothesize that VDR signaling regulates developmental trajectory of *A. limnaeus* embryos and orchestrates the expression of the complex phenotype associated with diapause.

Exogenous Vitamin D₃ Analogs Induce the Escape Trajectory Under Environmental Conditions That Favor Induction of Diapause. All analogs of vitamin D₃ were able to induce the escape trajectory under conditions that should favor diapause (20 °C to 25 °C), with 25-hydroxyvitamin D₃ [25(OH)D₃] and 1,25(OH)₂D₃ exhibiting potency in the picomolar range (Fig. 3). Surprisingly, 25(OH)D₃ is the more potent form of vitamin D₃. This may be due to higher expression of the 1,25(OH)₂D₃-24-hydroxylase enzyme in diapause trajectory embryos (Fig. 2) that inactivates 1,25(OH)₂D₃, and/or the need for local conversion to 1,25(OH)₂D₃ for maximum potency. Interestingly, 7-DHC was only able to induce the escape trajectory at 25 °C, which may indicate that 20 °C is below the threshold of thermal activity required to convert 7-DHC to vitamin D₃. These data, combined with the higher levels of 1,25(OH)₂D₃ in escape trajectory embryos, provide strong evidence of a biosynthetic pathway for synthesis of 1,25(OH)₂D₃ in embryos of *A. limnaeus* that promotes active development in a manner parallel to the action of dafachronic acids in *C. elegans*.

In humans, conversion of 7-DHC to vitamin D₃ requires photochemical conversion via UVB radiation and a heat-dependent isomerization (27). However, synthesis of 1,25(OH)₂D₃ is possible in a number of species without exposure to UVB, and blue light can induce production of vitamin D₃ in the skin of rainbow trout (28, 29). Further, in poikilotherms, thermal conversion of vitamin D₃ from previtamin D₃ is over a thousand times more efficient in cellular membranes than in solution (30), consistent with a role for the cellular microenvironment, or a possible enzymatic mechanism to facilitate conversion. Environmental activation of vitamin D₃ synthesis is consistent with laboratory data where short (24 h to 48 h) exposures to light and elevated temperature are known to induce the escape trajectory (SI Appendix, Fig. S5), and release embryos from dormancy in diapause (4). The potential for solar radiation and increased temperature to induce vitamin D₃ synthesis presents an opportunity for environmental factors to directly affect developmental trajectory.

Hormones That Promote Active Development in *C. elegans* Can Induce the Escape Trajectory in Embryos of *A. limnaeus*. Exposure of *A. limnaeus* embryos to Δ4-dafachronic acid induces the escape trajectory under conditions that favor entrance into diapause (Fig. 4). This effect is likely mediated through the VDR, given the strikingly similar developmental phenotypes observed between embryos treated with dafachronic acid and 1,25(OH)₂D₃. Further, we find no evidence that known agonists of closely related NRs (pregnane X, farnesoid X, and liver X) can induce the escape phenotype (SI Appendix, Table S2). Dafachronic acids are the major ligands for the *daf-12* NR in *C. elegans* that promote active development and prevent entrance into dauer dormancy (15). These data suggest a high degree of functional homology between *daf-12* and the vertebrate VDR, and a more general

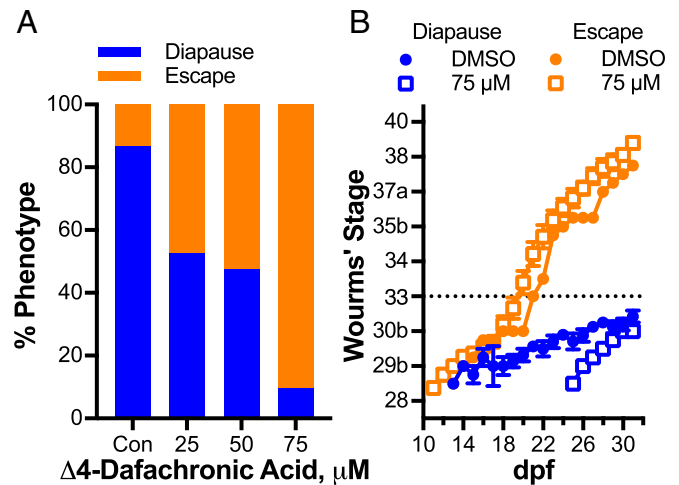


Fig. 4. Δ4-dafachronic acid induces the escape trajectory in *A. limnaeus* embryos. (A) Δ4-dafachronic acid causes embryos of *A. limnaeus* to follow the escape trajectory under conditions (25 °C) that should favor entrance into diapause. (B) Developmental rate and pattern of escape embryos induced by dafachronic acids are indistinguishable from normal escape embryos. However, embryos that follow the diapause trajectory despite treatment with dafachronic acids have a delayed development. Symbols are means ± SEM (*n* = 12 to 24). Dashed line at Wourms' stage 33 is diapause II. dpf, days postfertilization.

role for hormones derived from 7-DHC in the regulation of life history characters across a deep evolutionary divergence.

Blocking Synthesis of 1,25(OH)₂D₃ Induces Diapause in *A. limnaeus* and a Diapause-Like State in Zebrafish. Dafadine A, a potent inhibitor of the *daf-9* cytochrome P450 enzyme and its mammalian ortholog CYP27A1 (vitamin D₃ 25-hydroxylase), can block synthesis of 1,25(OH)₂D₃ (31). When *A. limnaeus* embryos are incubated at 30 °C in the presence of 10 μM to 25 μM dafadine A, most embryos enter diapause (Fig. 5). Thus, inhibition of 1,25(OH)₂D₃ synthesis results in embryos entering diapause under conditions that should result in active development. Importantly, this phenotype can be rescued by the addition of 100 pM 1,25(OH)₂D₃ (Fig. 5B). These data provide strong evidence that synthesis of 1,25(OH)₂D₃ controls developmental phenotype in embryos of *A. limnaeus*.

Surprisingly, incubation of zebrafish embryos in dafadine A resulted in a developmental and physiological arrest that is strikingly similar to diapause II in *A. limnaeus* (Fig. 6 and SI Appendix, Table S3). Recovery from dafadine exposure leads to a resumption of normal morphological and physiological development in most (60 to 100%) embryos (Fig. 6 and SI Appendix, Fig. S6). In contrast to embryos of *A. limnaeus*, this phenotype cannot be rescued by addition of up to 25 μM 1,25(OH)₂D₃. The reason for this difference is not known, but may be due to zebrafish expressing both VDR-A and B isoforms (32), while *A. limnaeus* expresses mainly the VDR-B isoform (Fig. 2). In fish, the two isoforms have been found to be functionally different in their response and sensitivity to 1,25(OH)₂D₃ (32, 33). It is also possible that dafadine A may interact directly with the VDR-A isoform and block the action of 1,25(OH)₂D₃, or induce expression of genes that deactivate 1,25(OH)₂D₃. Additional studies are needed to explore these possibilities. Despite species-specific differences, it appears that vitamin D₃ signaling promotes active vertebrate development and suggests that diapause-like states may be induced in other fishes through alteration of vitamin D₃ signaling.

VDR and Ciliary Signaling May Regulate Downstream Pathways That Define Developmental Phenotype. To explore the possible transcriptomic effects of VDR signaling, we interrogated the top hub genes associated

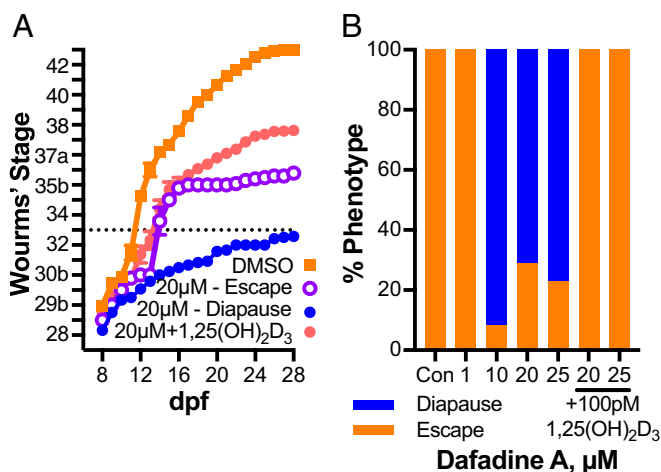


Fig. 5. Dafadine A induces diapause in *A. limnaeus* embryos. (A) Dafadine A causes embryos of *A. limnaeus* to enter diapause under conditions (30 °C) that should favor active development. (B) The effect of dafadine A on developmental trajectory can be reversed by addition of 100 pM 1,25(OH)₂D₃, although developmental rate is affected. Symbols are means ± SEM ($n = 12$ to 24). Dashed line at Wourms' stage 33 is diapause II. dpf, days postfertilization.

with the five WGCNA modules that were not preserved in the two trajectories and were correlated with embryonic stages after the temperature-sensitive window of commitment (identified as salmon, light green, tan, brown, and gray in Fig. 1A). Two of these modules can be directly linked to VDR signaling and contain a number of transcripts that are relevant to stress tolerance (salmon and brown), while two others (light green and gray) have hub genes that may play critical roles in the regulation of ciliary signaling (Dataset S3). For a more in-depth discussion of these modules and their potential importance in the determination of the diapause phenotype, please see *SI Appendix*.

Only a Small Number of Stress-Responsive Genes Are Up-Regulated in Diapause-Bound Embryos. Increased and sometimes extreme stress tolerance is an unexplained correlate of metabolic dormancy across all domains of life. Surprisingly, there is not an overall robust transcriptional “stress response” in embryos destined to enter diapause. In fact, for many stress-responsive genes, especially those encoding for heat shock chaperones and antioxidant proteins, expression is higher in the escape trajectory embryos (*SI Appendix, Fig. S7 C and F*). One exception to this pattern is a small cluster of heat shock proteins and a heat shock factor in the brown module that are strongly up-regulated in diapause-bound embryos (*SI Appendix, Fig. S7E*). There are also a small number of antioxidant genes that are much more abundant in diapause-bound embryos (*SI Appendix, Fig. S7C*). One clear relationship is a higher expression of genes involved in DNA repair and genome maintenance in diapause-bound embryos (*SI Appendix, Fig. S7A*). These expression patterns provide the basis for exploring the molecular underpinnings of stress tolerance associated with diapause.

The Potential Importance of IL-1 β in Regulating Developmental Phenotype. Cytokine signaling in early fish development has received very little attention, despite its ubiquitous expression in early zebrafish embryos (13). Please see *SI Appendix* for a brief discussion of a potential role for IL-1 β in regulating temperature-dependent phenotypes in *A. limnaeus*.

Discussion

Here we present evidence that VDR signaling regulates developmental trajectory associated with diapause in embryos of *A. limnaeus* in a manner that is consistent with daf-12 regulation of

dauer diapause in *C. elegans* (34). We report a role for vitamin D₃ signaling in the determination of life history decisions during vertebrate development, and describe a molecular signaling pathway that regulates developmental trajectory and entrance into metabolic dormancy. Perhaps the most significant implication of this work is the discovery of a conserved function for NRs in the regulation of metabolic dormancy in animals, and the potential to induce diapause-like states in species that do not naturally arrest development. Further, gene expression data in *Xenopus* and Atlantic salmon suggest an unexplored role for VDR in amphibian metamorphosis and salmonid smoltification (35, 36), which suggests a potential broader role for vitamin D₃ signaling in major life history transitions in vertebrates.

The VDR is a member of a family of NRs that are related to the ecdysone receptor in *Drosophila* and the daf-12 NR in *C. elegans* (37). The importance of these receptors in the regulation of life history decisions in nematodes and arthropods is clearly established, and evidence presented in this study extends this role to vertebrates. It is well understood that environmental induction of insulin/IGF1, TGF β , and cGMP signaling pathways converge on DAF-12 as the key regulator of developmental phenotype in *C. elegans* (34). In *Drosophila*, ecdysone signaling is known to integrate and transduce environmental signals that regulate adult diapause (38, 39). Interestingly, the synthesis of dafachronic acids, ecdysteroids, and vitamin D₃ all depend on 7-DHC as the initial precursor. This suggests that 7-DHC-dependent hormone synthesis could represent an environmental-specific hormonal signaling pathway across all animals, with hormonal chemistry and NR sequence evolving in a concerted response to each organism's natural environment.

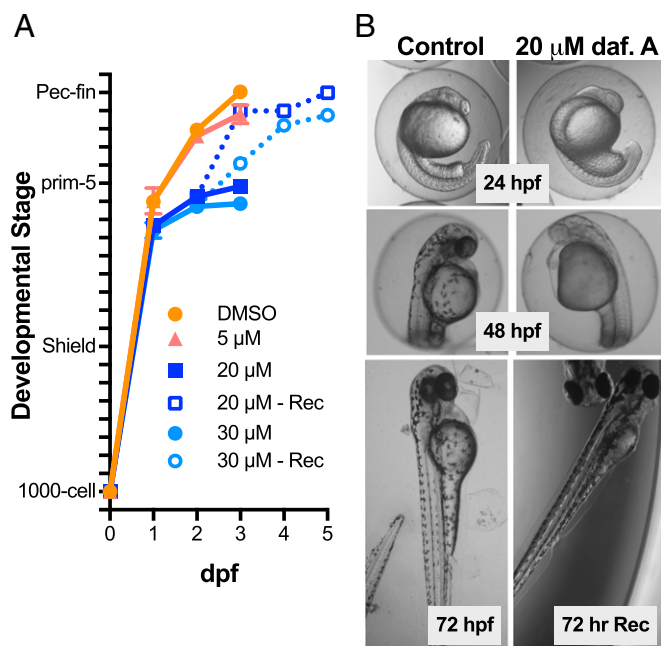


Fig. 6. Dafadine A induces a diapause-like state in zebrafish embryos. (A) Dafadine A arrests zebrafish development near the prim-5 stage of development and causes a phenotype that is strikingly similar to that of an *A. limnaeus* diapause embryo. Transfer of embryos treated for 48 h to dafadine-free media leads to a resumption of normal development. Symbols are means ± SEM ($n = 3$ groups of 20 embryos for normal development and 9 to 12 for recovery). (B) Images of zebrafish embryos exposed to control (1% DMSO) or 20 µM dafadine A. Zebrafish embryos were maintained at 28.5 °C and were initially exposed to dafadine A at the 1,000-cell stage. dpf, days postfertilization; hpf, hours postfertilization; Rec, recovery from dafadine A exposure. (Magnification, 40 \times .)

Materials and Methods

Experimental Design. Adult *A. limnaeus* were housed in the aquatic vertebrate facility at Portland State University (PSU) according to Institutional Animal Care and Use Committee (IACUC)-approved methods established for this species (PSU Protocol #33) (40, 41). Following collection, embryos were maintained for 4 d postfertilization (dpf) at 25 °C in darkness (40). Embryos were transferred to 20 °C and 30 °C at 100% epiboly. Embryos were flash frozen at seven stages as described in the caption of Fig. 1. Zebrafish embryos were obtained from the laboratory of Kim Brown at PSU and used according to PSU IACUC approved protocols (PSU Protocol #33) based on established optimal rearing conditions for zebrafish (42).

Transcriptome and WGCNA Analysis. RNA extraction, sequencing, and processing were performed as described previously (8). Gene counts were normalized as fragments per kilobase of transcript per million mapped reads (FPKM). Count matrices (all genes and sample libraries) were filtered for at least one normalized count across all replicates. Differential gene expression was determined between diapause and escape samples at each stage using DESeq2 in R Bioconductor (43). Significance was determined using a Benjamini–Hochberg false discovery rate (FDR) adjusted *P* value < 0.10. Coexpression

networks were constructed for each trajectory using the WGCNA R software package's default parameters (11). The accession identity of each NCBI biosample used in this study can be found in *SI Appendix, Table S1*.

Vitamin D₃ Analogs, Dafadine A Treatments, and Measurement of 1,25(OH)₂D₃. Killifish embryos were maintained at 25 °C with minimal light until 4 dpf, when embryos were transferred to experimental conditions. Chemicals were from Sigma-Aldrich (7-DHC, vitamin D₃, dafadine A), Selleck Chemicals [25(OH)D₃ and 1,25(OH)₂D₃] or Cayman Chemicals (Δ4-dafachronic acid). Embryos were monitored for development and staged under dim yellow light using a Leica model DMIRB inverted compound microscope (1, 42). Levels of free 1,25(OH)₂D₃ were measured using a calcitriol ELISA according to the manufacturer's instructions (Aviva Systems Biology).

ACKNOWLEDGMENTS. We thank Will Patterson for his help supporting the bioinformatics analyses and Andrew McCracken for collecting the data detailing the effects of light and heat on developmental trajectory presented in *SI Appendix, Fig. S4*. This work was supported by National Science Foundation Grant IOS 1354549 (to J.E.P.).

- Podrabsky JE, et al. (2017) Embryonic development of the annual killifish *Austrofundulus limnaeus*: An emerging model for ecological and evolutionary developmental biology research and instruction. *Dev Dyn* 246:779–801.
- Podrabsky J, Riggs C, Wagner J (2016) Tolerance of environmental stress. *Annual Fishes. Life History Strategy, Diversity, and Evolution*, eds Berois N, Garcia G, De Sá R (CRC Press, Boca Raton, FL), pp 159–184.
- Wourms JP (1972) The developmental biology of annual fishes. 3. Pre-embryonic and embryonic diapause of variable duration in the eggs of annual fishes. *J Exp Zool* 182: 389–414.
- Podrabsky JE, Hand SC (1999) The bioenergetics of embryonic diapause in an annual killifish, *austrofundulus limnaeus*. *J Exp Biol* 202:2567–2580.
- Chennault T, Podrabsky JE (2010) Aerobic and anaerobic capacities differ in embryos of the annual killifish *Austrofundulus limnaeus* that develop on alternate developmental trajectories. *J Exp Zool A Ecol Genet Physiol* 313:587–596.
- Podrabsky JE, Garrett IDF, Kohl ZF (2010) Alternative developmental pathways associated with diapause regulated by temperature and maternal influences in embryos of the annual killifish *Austrofundulus limnaeus*. *J Exp Biol* 213:3280–3288.
- Romney AL, Podrabsky JE (2017) Transcriptomic analysis of maternally provisioned cues for phenotypic plasticity in the annual killifish, *Austrofundulus limnaeus*. *EvoDevo* 8:6.
- Woll SC, Podrabsky JE (2017) Insulin-like growth factor signaling regulates developmental trajectory associated with diapause in embryos of the annual killifish *Austrofundulus limnaeus*. *J Exp Biol* 202:2777–2786.
- Martin KLM, Podrabsky JE (2017) Hit pause: Developmental arrest in annual killifishes and their close relatives. *Dev Dyn* 246:858–866.
- Wagner JT, et al. (2018) The genome of *Austrofundulus limnaeus* offers insights into extreme vertebrate stress tolerance and embryonic development. *BMC Genomics* 19:155.
- Langfelder P, Horvath S (2008) WGCNA: An R package for weighted correlation network analysis. *BMC Bioinformatics* 9:559.
- Langfelder P, Luo R, Oldham MC, Horvath S (2011) Is my network module preserved and reproducible? *PLoS Comput Biol* 7:e1001057.
- Sun W, Li Z, Zhang S (2014) Acute phase response in zebrafish embryo/larva with special emphasis on LPS-induced changes in expression pattern of acute phase protein genes. *J Mar Biol Assoc U K* 94:1569–1580.
- Fan J, Wojnar MM, Theodorakis M, Lang CH (1996) Regulation of insulin-like growth factor (IGF)-I mRNA and peptide and IGF-binding proteins by interleukin-1. *Am J Physiol* 270:R621–R629.
- Rottiers V, et al. (2006) Hormonal control of *C. elegans* dauer formation and life span by a Rieseke-like oxygenase. *Dev Cell* 10:473–482.
- Hu PJ (2007) Dauer. *WormBook*, 10.1895/wormbook.1.144.1.
- van Driel M, et al. (2006) Evidence for auto/paracrine actions of vitamin D in bone: α-hydroxylase expression and activity in human bone cells. *FASEB J* 20:2417–2419.
- Schuster I (2011) Cytochromes P450 are essential players in the vitamin D signaling system. *Biochim Biophys Acta* 1814:186–199.
- Kwon H-J (2016) Vitamin D receptor signaling is required for heart development in zebrafish embryo. *Biochem Biophys Res Commun* 470:575–578.
- Rosen CJ, et al. (2012) The nonskeletal effects of vitamin D: An endocrine society scientific statement. *Endocr Rev* 33:456–492.
- Henry HL, Norman AW (1978) Vitamin D: Two dihydroxylated metabolites are required for normal chicken egg hatchability. *Science* 201:835–837.
- Sunde ML, Turk CM, DeLuca HF (1978) The essentiality of vitamin D metabolites for embryonic chick development. *Science* 200:1067–1069.
- Howarth DL, et al. (2008) Paralogous vitamin D receptors in teleosts: Transition of nuclear receptor function. *Endocrinology* 149:2411–2422.
- Bethke A, Fielenbach N, Wang Z, Mangelsdorf DJ, Antebi A (2009) Nuclear hormone receptor regulation of microRNAs controls developmental progression. *Science* 324: 95–98.
- Carlberg C, Campbell MJ (2013) Vitamin D receptor signaling mechanisms: Integrated actions of a well-defined transcription factor. *Steroids* 78:127–136.
- Schröder M, Müller KM, Nayeri S, Kahler J-P, Carlberg C (1994) Vitamin D₃-thyroid hormone receptor heterodimer polarity directs ligand sensitivity of transactivation. *Nature* 370:382–386.
- Holick MF, et al. (1980) Photosynthesis of previtamin D₃ in human skin and the physiologic consequences. *Science* 210:203–205.
- Norman TC, Norman AW (1993) Consideration of chemical mechanisms for the non-photochemical production of vitamin D₃ in biological systems. *Bioorg Med Chem Lett* 3:1785–1788.
- Pierens SL, Fraser DR (2015) The origin and metabolism of vitamin D in rainbow trout. *J Steroid Biochem Mol Biol* 145:58–64.
- Holick MF, Tian XQ, Allen M (1995) Evolutionary importance for the membrane enhancement of the production of vitamin D₃ in the skin of poikilothermic animals. *Proc Natl Acad Sci USA* 92:3124–3126.
- Luciani GM, et al. (2011) Dafadine inhibits DAF-9 to promote dauer formation and longevity of *Caenorhabditis elegans*. *Nat Chem Biol* 7:891–893.
- Lin C-H, Su C-H, Tseng D-Y, Ding F-C, Hwang P-P (2012) Action of vitamin D and the receptor, VDRa, in calcium handling in zebrafish (*Danio rerio*). *PLoS One* 7:e45650.
- Kollitz EM, Hawkins MB, Whitfield GK, Kullman SW (2014) Functional diversification of vitamin D receptor paralogs in teleost fish after a whole genome duplication event. *Endocrinology* 155:4641–4654.
- Fielenbach N, Antebi A (2008) *C. elegans* dauer formation and the molecular basis of plasticity. *Genes Dev* 22:2149–2165.
- Li YC, Bergwitz C, Juppner H, Demay MB (1997) Cloning and characterization of the vitamin D receptor from *Xenopus laevis*. *Endocrinology* 138:2347–2353.
- Lock E-J, et al. (2007) The vitamin D receptor and its ligand 1α,25-dihydroxyvitamin D₃ in Atlantic salmon (*Salmo salar*). *J Endocrinol* 193:459–471.
- Whitfield GK, et al. (2003) Cloning of a functional vitamin D receptor from the lamprey (*Petromyzon marinus*), an ancient vertebrate lacking a calcified skeleton and teeth. *Endocrinology* 144:2704–2716.
- Emerson KJ, Bradshaw WE, Holzapfel CM (2009) Complications of complexity: Integrating environmental, genetic and hormonal control of insect diapause. *Trends Genet* 25:217–225.
- Saunders DS, Richard DS, Applebaum SW, Ma M, Gilbert LI (1990) Photoperiodic diapause in *Drosophila melanogaster* involves a block to the juvenile hormone regulation of ovarian maturation. *Gen Comp Endocrinol* 79:174–184.
- Podrabsky JE, Hrbeck T, Hand SC (1998) Physical and chemical characteristics of ephemeral pond habitats in the Maracaibo basin and Llanos region of Venezuela. *Hydrobiologia* 362:67–78.
- Podrabsky JE (1999) Husbandry of the annual killifish *Austrofundulus limnaeus* with special emphasis on the collection and rearing of embryos. *Environ Biol Fishes* 54: 421–431.
- Westerfield M (1995) *The Zebrafish Book: A Guide for the Laboratory Use of Zebrafish (Brachydanio rerio)* (M. Westerfield, Eugene, OR).
- Love MI, Huber W, Anders S (2014) Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol* 15:550.
- Shannon P, et al. (2003) Cytoscape: A software environment for integrated models of biomolecular interaction networks. *Genome Res* 13:2498–2504.
- Eisen MB, Spellman PT, Brown PO, Botstein D (1998) Cluster analysis and display of genome-wide expression patterns. *Proc Natl Acad Sci USA* 95:14863–14868.
- de Hoon MJL, Imoto S, Nolan J, Miyano S (2004) Open source clustering software. *Bioinformatics* 20:1453–1454.
- Saldanha AJ (2004) Java Treeview—Extensible visualization of microarray data. *Bioinformatics* 20:3246–3248.